

**Bioaccumulation and effects of metals bound to sediments collected from Gulf of
Cádiz (SW Spain) using the polychaete, *Arenicola marina***

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1 **Introduction**

2

3 Sediments are widely recognized and employed in assessing the impact of contaminants
4 on aquatic systems (Bryan and Langston 1992) as chemicals normally have
5 considerable higher concentrations in sediments than in the water column. Given that
6 aquatic sediments can act as reservoir for persistent contaminants (e.g. trace metals),
7 organisms which live in and feed on sediment can accumulate considerable amount of
8 contaminants. Thus, to assess the adverse effects of pollutants on the aquatic
9 sedimentary environment, several authors have proposed the use of sediment toxicity
10 tests (Chapman and Wang 2001). These kinds of tests aim to study the relationships
11 between the sediment concentration of chemicals and any adverse biological effects on
12 the biota resulting from exposure to these chemicals. Therefore, the choice of test
13 organisms is essential for providing appropriate information of the hazards of chemical
14 stressors (Chapman et al. 2002). Polychaetes, in particular *A. marina* have been used as
15 biomonitors of littoral and estuarine contamination (Packer et al. 1980; Bat 1998;
16 OSPAR 1995; Casado-Martínez et al. 2008; Morales-Caselles et al. 2008). Lugworms
17 are abundant, tolerant to a wide range of environmental factors and ecologically
18 important, moreover they live in direct contact with the sediment.
19 The majority of studies on metal availability in sediments are based on toxicity,
20 however bioaccumulation may be a more accurate endpoint for assessing bioavailability
21 in risk assessment (Ankley et al. 1996). Trace metals may be bioavailable even if
22 toxicity is not observed since toxicity depends on various factors such as the sensitivity
23 of organism, length of exposure period etc. Biological adverse effects of pollutants may
24 also be manifested at the biochemical level in organisms or at higher levels of
25 organisation and responses are measurable. Biomarkers have been incorporated into

26 environmental toxicology research for decades, but they are still seldom used in a day to
27 day management of the environment (Handy et al. 2003). The biomarker based on the
28 measurement of metallothionein-like protein (MTLP) concentration in biological tissues
29 represents a detoxification role for structures which have been reversibly impaired by
30 inappropriate metal binding (Martín-Díaz et al. 2007). They have been considered as
31 useful and suitable biomarkers of metal exposure in aquatic organisms (see review by
32 Amiard et al. 2006).

33 The aim of this work was to assess the bioavailability of metals bound to sediments
34 collected from twelve sampling sites along the Gulf of Cádiz, characterized with
35 different types and degrees of metal contaminations. In order to assess the defence
36 mechanisms against the accumulated metals, the induction of a typical biomarker,
37 metallothionein-like proteins was determined in tissues sampled after 7 and 14 days of
38 exposure. Data were integrated by multivariate analysis to correlate the concentrations
39 of contaminants and biological responses in the biota.

40

41 **Materials and Methods**

42

43 Study area, sample collection and characterization

44 Sediments were collected from twelve sampling sites along the southwest coast of Spain
45 (Table 1). Guadiana (sample 1, near the international bridge between Spain and Portugal
46 and sample 2 in the mouth of the river) and Huelva estuaries (sample 3 is more
47 influenced by the rivers Tinto and Odiel, and sample 4 in the Pedro Santo Channel) are
48 influenced by the surrounding mining area, moreover an industrial complex is located in
49 the proximity of Ría of Huelva. Sampling points in the Guadalquivir estuary (5 and 6)
50 are impacted by maritime transport and urban activities, as well as samples from the

51 Bay of Cádiz (samples 7 and 8), while sample 9 is a comparatively clean area
52 considered suitable as negative toxicity control according to previous studies (Riba et al.
53 2004a). The Bay of Algeciras (sample 10 and 12 in the mouth of Guadarranque River,
54 near chemical processing plants; sample 11 in the mouth of river Palmones) is
55 considered as an important harbour and receives considerable contaminant input derived
56 from several industrial and commercial shipping activities.

57 Sediments were collected with a 0.025 m² Van Veen grab from approximately the top
58 20 cm of the sediments. Samples were transferred to the laboratory where they were
59 homogenized, sieved through a 2 mm mesh and stored in dark at 4 °C no longer than 2
60 weeks before the start of testing. Total organic carbon (TOC) was determined using the
61 El Rayis (1985) modification based on an acidification of the sediment sample, and
62 grain size distribution was analysed by using a laser particle size Frisch (model
63 Analysette 22) following the method reported by Riba et al. (2003).

64

65 Toxicity test

66 The test organisms, *Arenicola marina* were collected in the Cantabrian coast (North
67 Spain) by hand digging and were transported to the laboratory in cool boxes containing
68 clean seawater. Worms were transferred to 20 L tanks containing clean sediment (5 cm
69 depth) and seawater and acclimated to laboratory conditions for a week prior to the test
70 (17±1°C, salinity 38). Test organisms were placed in tanks containing only seawater to
71 depurate the digestive tract 24 h prior to tests. The bioassay was performed in duplicate.
72 Small tanks were filled with approximately 1 kg of sediments and 5 cm overlying clean
73 seawater provided with gentle aeration. After the acclimation period 7 organisms were
74 placed in every tank and after 24 h unburied worms were replaced. During the 14 days
75 of exposure, the overlying water was replaced every two days and physical parameters

76 (temperature, pH, salinity, dissolved oxygen concentration) were measured in each tank
77 on the same day. Lugworms were sampled on day 0 to obtain the initial concentrations
78 of tissues. After 7 and 14 days 4 individuals from each station were collected and 2 of
79 them were immediately frozen at -80 °C (MTLP measurement), while the rest were
80 placed for 24 h in clean seawater for depuration and stored at -20°C until trace metal
81 analysis. The mortality was also recorded during the experimental period.

82

83 Trace metal analyses

84 The whole tissues of lugworms were lyophilised and digested in concentrated HNO₃
85 and H₂O₂ (Suprapur, Merck) at 95°C (García-Luque et al. 2007), while sediment
86 samples after freeze-drying were extracted with 1 N HCl to provide more information
87 about the bioavailability of metals (Luoma and Bryan 1982). In both cases, blanks were
88 prepared simultaneously in the same way as the sample. Concentrations of Cd, Co, Cu,
89 Ni, Pb and Zn in the acid digests were determined with an inductively coupled plasma-
90 optical emission spectrometer (Optima 2000 DV, Perkin Elmer). The accuracy of the
91 analyses was checked by digesting certified reference material (TORT-2, Lobster
92 Hepatopancreas, National Research Council, Canada) and was considered satisfactory
93 (Table 2). Data are expressed as µg g⁻¹ dry weight.

94

95 MTLP induction

96 Tissue samples were homogenized with an ultraturrax homogenizer with Trizma-
97 HCl/Trizma-Base 0.1M (pH 7.6) buffer (at a ratio of 1:3) on ice (4°C) and centrifuged at
98 30 000g for 2 h at 4°C. The supernatant (cytosol) was separated from the pellet and 0.1
99 mL of supernatant was added to 0.9 mL of NaCl (0.9%), heated at 95°C for 4 min and
100 centrifuged at 10 000g for 15 min at 4°C. Metallothionein-like protein concentrations

101 were determined in the supernatant by Anodic Stripping Voltametry (ASV), according
102 to the procedure described by Olafson and Olson (1991), using purified rabbit
103 metallothionein (Sigma-Aldrich, MT I-II). The total protein concentration was
104 determined with a bicinchoninic acid protein assay (Smith et al. 1985).

105

106 **Results and Discussion**

107

108 Chemical characteristics of sediments

109 The concentrations of trace metals (Cu, Zn, Cd, Ni, Co and Pb) presented in the twelve
110 sediments are shown in Table 3. Among all stations the negative control (sample 9) and
111 sample 5 from the Guadalquivir estuary showed the lowest metal concentrations in
112 sediments, whereas sediments from sample sites from Huelva estuary and Bay of
113 Algeciras (samples 3, 4, 6 and 10) contained greater amount of trace metals. For
114 instance, the highest concentrations of Cd, Zn, Cu and Pb were measured in sample 3.
115 Total organic carbon content and grain size most probably control metal concentrations
116 in sediments. The proportion of fines ranged between 30 and 90%, except samples 5
117 and 9 with a proportion of fines (referring to the proportion of sediment $< 63\mu\text{m}$) $<$
118 10%. Similarly, the TOC varied between 0.10 and 3%, except samples 5 and 9, which
119 had TOC values $< 0.10\%$. The correlation between the proportion of fines and TOC
120 concentrations in sediments was statistically significant ($p < 0.05$). It has to be mentioned
121 that independently of the exclusion of the sandy sediment samples, this relationship
122 behaved similar.

123

124 Biological responses in *Arenicola marina*

125 Organisms exposed to the sediment collected from the proximity of chemical processing
126 plants in the Bay of Algeciras (sample 10) showed the highest accumulated mortality at
127 the end of the exposure (66.6%), followed by those in Huelva estuary (sample 3) and
128 the two sampling points in the Guadalquivir estuary (sample 5 and 6). Toxic responses
129 measured in sediments from the mouth of Guadiana estuary (sample 2), Huelva estuary
130 (sample 4) and mouth of river Palmones (sample 11) were lower, while the rest of the
131 stations did not cause mortality.

132 The rate of increase in metal concentrations (K_{metal}) from days 0 to 14 in *A. marina* was
133 described by a linear kinetic approach (Martín-Díaz et al. 2007) and was fitted with the
134 results as follows:

$$135 \ln [M] = \ln [C_0] + K_t$$

136 where C_0 is the concentration of metal or MTLP at $t = 0$; M is the concentration of
137 metal or MTLP at time t minus at the initial time (0); K is the constant rate of increase.

138 The positive K values obtained from these equations demonstrated that lugworms had
139 potential to accumulate trace metals during the exposure time. In general, trace metal
140 concentrations were higher in specimens sampled from the contaminated sites. On day
141 14, elevated concentrations of Zn, Cu, Ni and Co were recorded in individuals from the
142 Bay of Algeciras (sample 10), as well as Cu, Zn in lugworms exposed to sediments
143 from Huelva estuary (samples 3 and 4). Part of the samples were accidentally lost
144 during the analysis, therefore no data for K_{Pb} are available.

145 The rate of metallothionein-like protein increase (K_{MT}) demonstrated the induction of
146 these proteins in organisms along the 14 days of exposure. Lowest MTLP synthesis was
147 observed in lugworms from samples 2 and 9, whereas increased concentrations were
148 elevated in various samples, such as 3, 5, 10 and 11.

149

150 Multivariate analysis

151 In order to observe the relationships between the chemical concentrations in sediments
152 and biological effects on test organisms, multivariate analysis was applied. The initial
153 data matrix contains 12 cases, the twelve stations and 15 variables, the geochemical
154 characteristics (fines and total organic carbon), the metal concentrations in sediments
155 (Cd, Zn, Ni, Co, Cu and Pb), the toxicity measured as mortality, the rate of increase of
156 MT concentrations (K_{MT}) and the rate of increase of bioaccumulation of metals in
157 tissues (K_{Cd} , K_{Zn} , K_{Ni} , K_{Co} , K_{Cu}). The factor analysis with varimax normalized permitted
158 after the reduction of 15 variables to three principal factors, which explains 79.14 % of
159 the total variance (Table 4).

160 The first principal factor (F1) was predominant and accounted for 41.75% of the
161 variance. This factor combines concentrations of Cd, Zn, Cu and Pb in sediment and
162 shows relationship with the induction of MTs (K_{MT}) and with the increase in
163 bioaccumulation of Zn and Cu (K_{Zn} , K_{Cu}). The second principal factor (F2) accounts for
164 21.35% of the total variance and has positive loadings for the sediment concentrations
165 of Ni and Co, the fine particles and total organic carbon of sediments and associates
166 with toxicity to *A. marina*. The third principal factor (F3) accounts for 16.04% of total
167 variance and combines the bioaccumulation of Cd, Zn, Ni, Co. This factor suggests that
168 the bioaccumulation of these trace metals was not associated neither with the biological
169 responses that we have studied nor with the concentrations of these metals measured in
170 sediments.

171 Estimated factor scores are to explain the prevalence of every component for each
172 station and used to confirm the factor descriptions (Figure 1).

173 The first factor (F1), which is representative of the contaminants (Cd, Zn, Cu, Pb) in
174 sediment, the bioaccumulation of Zn and Cu and the MTLP induction in tissues, has

175 positive loadings in samples 3 and 4 from the Huelva estuary. Both samples are
176 characterized by high concentrations of trace metals due to mining and industrial
177 activities (Riba et al. 2004a). The elevated concentrations of Zn and Cu measured in
178 tissues of *A.marina* according to F1 did not cause toxicity. The most likely explanation
179 could be that Cu and Zn are considered as strong inducers of MTLPs, as these proteins
180 can bind them via the sulfhydryl group of cysteine residues. Therefore MTLPs were
181 probably involved in protection against trace metal toxicity. The induction of MTLPs in
182 polychaetes after metal exposure has been confirmed by several authors. In the Cd-
183 exposed *Neanthes arenaceodentata* an increase in the metal concentration associated
184 with MTLPs was observed by Jenkins and Mason (1988). Ng et al. (2008) found no
185 increase in MTLP concentrations, but higher MTLP turnover (synthesis and breakdown)
186 in the nereid polychaete, *Perinereis aibuhitensis* after Cd pre-exposure. These findings
187 indicate that measuring the MTLP concentration alone can be misleading. The synthesis
188 of MTLPs was observed in *Eurythoe complanata* after exposure to Cu or Zn (Marcano
189 et al. 1996). However these authors reported no increase in the concentrations of
190 MTLPs in the combined Cu-Zn treatments, suggesting the possible interaction between
191 the metals. Trace metals in sediments from the Huelva estuary also caused MTLP
192 induction in female shore crabs, *Carcinus maenas* (Martín-Díaz et al. 2009) after 28
193 days of exposure. Numerous studies support the idea of using MTLP concentrations in
194 organisms as biomarkers in environmental monitoring programme. It has to be
195 mentioned that the expression of MTLPs can be influenced by various natural factors
196 that affect the metal accumulation, such as salinity (Leung et al. 2002) or temperature
197 (Serafim et al. 2002). In some species the presence of MTLPs has been confirmed but
198 their levels were not correlated to the local metal bioavailability due to the ability to
199 regulate their metal burden (Mouneyrac et al. 2003; Poirier et al. 2006). Thus, additional

200 research on the relationship between metal toxicity and MTLP induction in this species
201 is required before their routine use in biomonitoring programs. Conversely, the Ni and
202 Co sediment concentrations, which were comprised in F2 and showed positive factor
203 scores in sample 3, 6, 10 and 11 (Figure 1) did not cause MTLP induction, but related
204 to high mortality to *A. marina*. F2 is mainly prevalent in sample 10, collected from the
205 Bay of Algeciras, continuously impacted by industrial effluents and maritime activities.
206 Morales-Caselles et al. (2007) previously reported high toxicity to *Corophium volutator*
207 exposed to sediment collected from the same area due to Ni and Co contaminants
208 associated with PAHs in oil spills. *A. marina* was chosen as an organism known to be
209 sensitive to hydrocarbons, but no measurements of these contaminants were included in
210 the present work. These two metals obviously could originate from any other sources,
211 however Riba et al. (2004b) reported hydrocarbon contamination in sediments from the
212 Huelva estuary (sample 3). Cesar et al. (2007) found that contaminations by PAHs and
213 Ni were closely related to amphipods (*C. volutator*) mortality in the Bay of Algeciras as
214 well as in the Huelva estuary. The characterization of sediments can provide some
215 explanations for the metal accumulation in the organisms. Usually the trace metal
216 concentrations increase with decreasing grain size of the sediment. In the case of fine
217 sediments, the surface binding sites per unit mass of the particle increase. Organic
218 matter can contain functional groups that form complexes with metals and decrease
219 their bioavailability to organisms. Negative correlations between bioavailability of some
220 metals to *A. marina* and the proportion of fines and total organic material have been
221 considered by Casado-Martínez (2006). It has been shown that deposit-feeding
222 invertebrates can adjust their rates of ingestion according to the quality of the sediment
223 (Cammen 1980). Thus, lower organic matter content may lead to an increase in
224 ingestion rate to satisfy their nutrient requirement. In the present study, lowest TOC

225 were recorded for two samples (5 and 9), in which low concentrations of metals were
226 measured, and as a consequence the bioaccumulation in lugworms exposed to these two
227 sediments was obviously low. Although higher TOC values were related to greater
228 amounts of accumulated metals by organisms (e.g. samples 3 and 10), these metals
229 generally are associated with organic materials in the sediments, that lugworms feed
230 preferentially.

231 The third factor demonstrates the bioaccumulation of Cd, Zn, Ni, Co in the tissues
232 without toxic or sublethal effect on the test organisms. The capacity of lugworms to
233 accumulate trace metals from sediments has been addressed by several authors (Packer
234 et al. 1980; Bat 1998; Bat and Raffaelli 1998; Casado-Martínez et al. 2008). Time
235 course of metal uptake was also revealed in three body compartments (body wall,
236 intestine and blood) of *A. marina* after 25 days of copper sediment exposure (Everaarts
237 1986).

238

239 **Conclusions**

240

241 The results of this survey confirm that MTLPs induction in *A. marina* was related to
242 increased sediment metal concentrations in samples collected from the Huelva area. The
243 rates of increase in MTLP and accumulated metal concentrations along the exposure
244 time seemed to be good and suitable tools for evaluation of metal bioavailability in
245 sediments. Results suggest that MTLP induction in tissues played an important
246 protective role, therefore lower toxicity was detected. In our study, the test organism,
247 *Arenicola marina* due to its capacity to accumulate metals from sediment was found as
248 a possible indicator species for sediment quality assessment. The use of multivariate

249 analysis appeared to be a useful tool to link the chemical concentrations and biological
250 responses.

251

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253

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257

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Figure captions

Figure 1. Factor loadings for the three principal factors in each of the twelve cases. The factor scores quantify the prevalence of every factor for each station and are used to establish the description of each factor.